

## Short Communication

# Red wine prevents the postprandial increase in plasma cholesterol oxidation products: a pilot study

F. Natella<sup>1\*</sup>, A. Macone<sup>2</sup>, A. Ramberti<sup>1</sup>, M. Forte<sup>1</sup>, F. Mattivi<sup>3</sup>, R. M. Matarese<sup>2</sup> and C. Scaccini<sup>1</sup>

<sup>1</sup>National Research Institute on Food and Nutrition, Via Ardeatina, 546, 00178 Rome, Italy

<sup>2</sup>Department of Biochemical Sciences, University La Sapienza, Rome, Italy

<sup>3</sup>Fondazione Edmund Mach, IASMA Research and Innovation Centre, Via E. Mach 1, 38010 San Michele all'Adige, Italy

(Received 24 June 2010 – Revised 23 November 2010 – Accepted 29 November 2010 – First published online 4 February 2011)

### Abstract

Moderate wine consumption has been shown to lower cardiovascular risk. One of the mechanisms could involve the control of postprandial hyperlipaemia, a well-defined risk factor for atherosclerosis, reasonably by reducing the absorption of lipid oxidised species from the meal. The objective of the present study was to investigate whether wine consumption with the meal is able to reduce the postprandial increase in plasma lipid hydroperoxides and cholesterol oxidation products, in human subjects. In two different study sessions, twelve healthy volunteers consumed the same test meal rich in oxidised and oxidisable lipids (a double cheeseburger), with 300 ml of water (control) or with 300 ml of red wine (wine). The postprandial plasma concentration of cholesterol oxidation products was measured by GC–MS. The control meal induced a significant increase in the plasma concentration of lipid hydroperoxides and of two cholesterol oxidation products, 7- $\beta$ -hydroxycholesterol and 7-ketocholesterol. The postprandial increase in lipid hydroperoxides and cholesterol oxidation products was fully prevented by wine when consumed with the meal. In conclusion, the present study provides evidence that consumption of wine with the meal could prevent the postprandial increase in plasma cholesterol oxidation products.

**Key words:** Red wine: Postprandial oxidative stress: Oxysterols: Human studies

Epidemiological studies have indicated that wine can be considered protective against CVD development when its moderate consumption is inserted in a correct lifestyle<sup>(1)</sup>, including the ‘instructions to drink/use’, i.e. ‘to be taken with meals’. A number of experimental studies have suggested that red wine compounds, especially polyphenols, might play a role in preventing the development and progression of atherosclerosis, acting through different pathways that include inhibition of lipid peroxidation, metal chelation, free-radical scavenging, inhibition of platelet aggregation, anti-inflammatory and oestrogenic activity, improvement of endothelial function, lowering of blood pressure and modulation of lipoprotein metabolism<sup>(2)</sup>. The attenuation of postprandial oxidative stress could be one of the mechanisms explaining the protective action of wine phenols<sup>(3,4)</sup>. In fact, the absorption of pro-oxidant/oxidised species with the meal can induce physiological events, such as the formation of mildly oxidised lipoprotein<sup>(5)</sup> or endothelial

dysfunction<sup>(6)</sup>, and inflammatory responses<sup>(7)</sup>, all events linked to the development of CVD.

There is evidence that oxysterols are angiotoxic and could cause atherosclerosis<sup>(8)</sup>. Animal studies have shown that the addition of oxidised cholesterol to the diet increases atherosclerosis<sup>(9)</sup>, and epidemiological studies have shown an association between plasma oxysterols and CVD<sup>(10)</sup>. Oxysterols have also shown to possess mutagenic and carcinogenic effects in both *in vivo* and *in vitro* studies<sup>(11)</sup>.

The typical Western diet contains substantial quantities of oxidised cholesterol, and the mean dietary intake has been estimated in mg/d per person<sup>(12)</sup>.

In view of the health implications of oxysterol absorption from food, we investigated, in a pilot study, the possibility that wine consumption with a meal influences the postprandial increase in plasma lipid hydroperoxides and oxysterols in humans.

**Abbreviation:** TMS, trimethylsilyl.

\*Corresponding author: Dr F. Natella, fax +39 6 51494550, email [natella@inran.it](mailto:natella@inran.it)

## Subjects and methods

### Subjects and study design

A total of twelve volunteers (six males and six females, age 24–35 years) participated in a cross-over study. Subjects, free from known diseases, were asked to keep their diet as constant as possible during the study period, and none of them was taking any drugs or vitamin supplement. The present study was conducted according to the guidelines laid down in the Declaration of Helsinki, and all procedures involving human subjects were approved by the Ethical Committee of the National Institute for Food and Nutrition Research. Verbal informed consent was obtained from all subjects; verbal consent was witnessed and formally recorded. Subjects ate the same test meal in two different sessions (2 weeks apart) after a 10–12 h fasting interval. The test meal, a double cheeseburger, was eaten with 300 ml of water (control) or with 300 ml of red wine (Teroldego Rotaliano, Foradori, 2004). The cheeseburger weighed approximately 200 g and contained 25.7 g of protein, 25.9 g of lipid (10.5 g SFA, 8.6 g MUFA and 0.8 g PUFA), 34.3 g of carbohydrate and 83 mg of cholesterol (US Department of Agriculture food composition table). The alcoholic grade of wine was 13.3 %, with pH 3.71, total SO<sub>2</sub> 66 mg/l, and its phenolic content was assessed as reported by Canali *et al.*<sup>(13)</sup>, with the exception of flavanols, which were estimated by LC–MS according to Mattivi *et al.*<sup>(14)</sup>.

### Plasma and meal analyses

Blood was collected before (time 0) and 1 and 3 h after the meal. Venous blood samples were collected into vacutainers containing opportune anticoagulants. Plasma samples were separated by centrifugation and stored at –80°C until analysis. Plasma total cholesterol, TAG and alcohol were measured by commercial kits (Futura System Srl, Formello, Roma, Italy; Sigma, St Louis, MO, USA). Plasma samples for the determination of oxysterols were stored at –80°C, after the addition of butylated hydroxytoluene (50 µg/ml), and analysed within 2 weeks. Total lipid hydroperoxides were measured in plasma by the ferrous ion oxidation xylene orange-2 assay, as described by Nourooz-Zadeh<sup>(15)</sup>.

Cheeseburger samples were analysed with the same methods described for plasma after homogenisation and extraction with chloroform–methanol<sup>(16)</sup>.

### Oxysterol measurement

The following four different oxysterols were measured in both the meal and plasma: 7-ketocholesterol (7-Keto-C), 5α,6α-epoxycholesterol, 5β,6β-epoxycholesterol and 7β-hydroxycholesterol by GC–MS<sup>(17)</sup>.

The four oxysterols were selected because they are the most abundant in food and efficiently absorbed<sup>(18)</sup>.

Briefly, plasma samples (200 µl) were added with 1 µg of the internal standard (19-hydroxycholesterol). Saponification was carried out under N<sub>2</sub> flow at 60°C for 90 min with 1 ml of 1 M-NaOH ethanolic solution. Samples were then extracted with cyclohexane, and the resulting organic layer was

evaporated to dryness under N<sub>2</sub>. Then, they were resuspended in 1 ml hexane and applied to solid-phase extraction (Supelclean Lc-Si cartridge; Sigma)<sup>(19)</sup>. The oxysterol fraction was dried under N<sub>2</sub> and derivatised with 70 µl of the Sylon BTZ kit (at room temperature for 45 min). GC–MS analyses were performed on an Agilent 6850A gas chromatograph coupled to a 5973N quadrupole mass-selective detector (Agilent Technologies, Palo Alto, CA, USA). Gas chromatographic separations were carried out on an Agilent HP-5MS fused silica capillary column (inner diameter 30 m × 0.25 mm and film thickness 0.25 µm). The injection mode was splitless at a temperature of 280°C. The column temperature programme was as follows: 160°C (1 min) to 280°C at a rate of 20°C/min and held for 15 min. The carrier gas was He at a constant flow of 1.0 ml/min. The spectra were obtained in electron impact mode at 70 eV ionisation energy; ion source temperature was 280°C and ion source vacuum was 10<sup>–5</sup> Torr (1.3 × 10<sup>–3</sup> Pa). Analyses were performed both in total ion current and selected-ion monitoring modes. Selected-ion monitoring analyses were carried out by selecting the following representative ions: *m/z* 353 for the 19-OH-C trimethylsilyl (TMS) derivative; *m/z* 456 for the 7β-hydroxycholesterol TMS derivative; *m/z* 474 for the 5β,6β-epoxycholesterol TMS derivative; *m/z* 474 for the 5α,6α-epoxycholesterol TMS derivative; *m/z* 472 for the 7-Keto-C TMS derivative.

### Statistical analysis

Data are presented as means and standard deviations. Statistical analysis was carried out using repeated-measures ANOVA, followed by Tukey's test for multiple comparisons. Analyses were performed with KaleidaGraph software (version 3.6; Synergy Software, Reading, PA, USA). *P* values < 0.05 were considered statistically significant.

## Results

### Wine composition

Total polyphenols (1871 mg/l, as catechin equivalents) were in the typical range for the variety. The concentration of total proanthocyanidins was 167.7 mg/l. The total administered dose of the major phenolics was calculated from the concentration in wine measured by HPLC at the time of the experiment. The wine had a quite high content of free anthocyanins, and the total administered dose was of 304.1 µmol. Hydroxycinnamates (85.2 µmol administered) consisted mainly of *trans*-caftaric acid, coumaric acid and *trans*-coumaric acid, with minor amounts of ferulic acid and grape reaction product (i.e. *trans*-2-S-glutathionyl-caftaric acid). Free flavanols (total of 82.9 µmol) consisted of epigallocatechin, (+)-catechin, epicatechin and gallocatechin. Myricetin was by far the main flavonol (20.5 µmol of total flavonols). Other minor phenolics were tyrosol (29.8 µmol) and the four monomers of resveratrol (for a total of 5.4 µmol).

In summary, the single dose of Teroldego wine provided 561 mg of phenolics (which is approximately in the millimolar level, assuming an average molecular weight of 500).

### Lipid hydroperoxides and oxysterols in the test meal

The lipid hydroperoxide content of the test meal was 237 (SD 36)  $\mu\text{mol}$  of  $\text{H}_2\text{O}_2$  equivalents.

As for oxysterols, the test meal contained 498 (SD 147)  $\mu\text{g}$  of 7-ketocholesterol, 138 (SD 48)  $\mu\text{g}$  of 5 $\alpha$ ,6 $\alpha$ -epoxycholesterol, 91 (SD 6)  $\mu\text{g}$  of 7 $\beta$ -hydroxycholesterol and 70 (SD 6)  $\mu\text{g}$  of 5 $\beta$ ,6 $\beta$ -epoxycholesterol.

When expressed per g of the test meal, total oxysterols were 3.9 (SD 1.1)  $\mu\text{g/g}$ , and this value is in accordance with literature data. van de Bovenkamp *et al.*<sup>(12)</sup> reported 3.6  $\mu\text{g}$  of total oxysterols/g of a cooked mixed Dutch diet, while Baggio *et al.*<sup>(20)</sup> and Rodriguez-Estrada *et al.*<sup>(21)</sup> reported a concentration of about 2  $\mu\text{g/g}$  of hamburger.

### Effect of the control and wine meals on plasma lipids, lipid hydroperoxides and oxysterols

Plasma concentrations of total cholesterol, TAG and alcohol, before and after the control and wine meals, are shown in Table 1. As expected, there was an increase in plasma TAG after the consumption of both meals, while ethanol, as expected, increased significantly only after the wine meal. Cholesterol concentration did not change significantly after both meals.

As shown in Fig. 1, the control meal induced a significant increase in total plasma lipid hydroperoxides. On the contrary, the wine meal not only prevented this increase, but also reverted it, inducing a significant decrease in plasma lipid hydroperoxides.

Fig. 1 shows also the effect of the meal on plasma oxysterols. The control meal induced a significant increase in 7 $\beta$ -hydroxycholesterol and 7-ketocholesterol concentrations. This increase was statistically significant 1 h after the consumption of the meal. The postprandial increase in these two oxysterols was fully prevented when wine was consumed with the meal. Indeed, wine consumption induced a significant decrease in 7 $\beta$ -hydroxycholesterol. Both 5 $\alpha$ ,6 $\alpha$ -epoxycholesterol and 5 $\beta$ ,6 $\beta$ -epoxycholesterol showed the same trend as observed for 7 $\beta$ -hydroxycholesterol and 7-ketocholesterol, even if their postprandial changes (after both the control and wine meals) were not statistically significant.

An estimation based on the subjects' volume of plasma (55% of volume of blood, calculated individually as 7% of their body weight<sup>(22)</sup>) indicates that total plasma oxysterols (the sum of the measured four oxysterols)

represented 105 (SD 29) and 95 (SD 26)% of the ingested dose 1 and 3 h after the control meal, respectively. As evident from Fig. 1, after the wine meal, total plasma oxysterols decreased below the baseline value ( $-55$  (SD 28) and  $-31$  (SD 41)% of the ingested dose, at 1 and 3 h, respectively).

### Discussion

Some authors suggest that the absorption from meals of the products of lipid oxidation could be, at least partially, the link between postprandial lipaemia and atherosclerosis<sup>(23)</sup>.

Oxysterols are a common component of the Western diet, and their presence is striking in fast food and processed food. Studies in both humans<sup>(18)</sup> and animals<sup>(24)</sup> have demonstrated that oxysterols are absorbed by the small intestine, transported in plasma by chylomicrons and incorporated into lipoprotein. As oxysterols possess several proatherogenic activities<sup>(8)</sup>, a delayed clearance of these compounds from the circulation could be harmful.

Although oxysterols are principally derived from dietary sources, circulating oxysterols may be produced enzymatically at the intracellular level and/or from lipoprotein oxidation into the circulation<sup>(25)</sup>, or by free radical-catalysed oxidation of cholesterol during digestion, both at gastric<sup>(4,26)</sup> and intestinal levels<sup>(27)</sup>. Even if oxysterols have a faster plasma clearance than 'normal' cholesterol, the level of oxysterols in plasma can remain elevated for more than 6–8 h after a meal<sup>(18)</sup>. Thus, the frequent consumption of foods rich in oxysterols can result in a continuous exposure during most of the day.

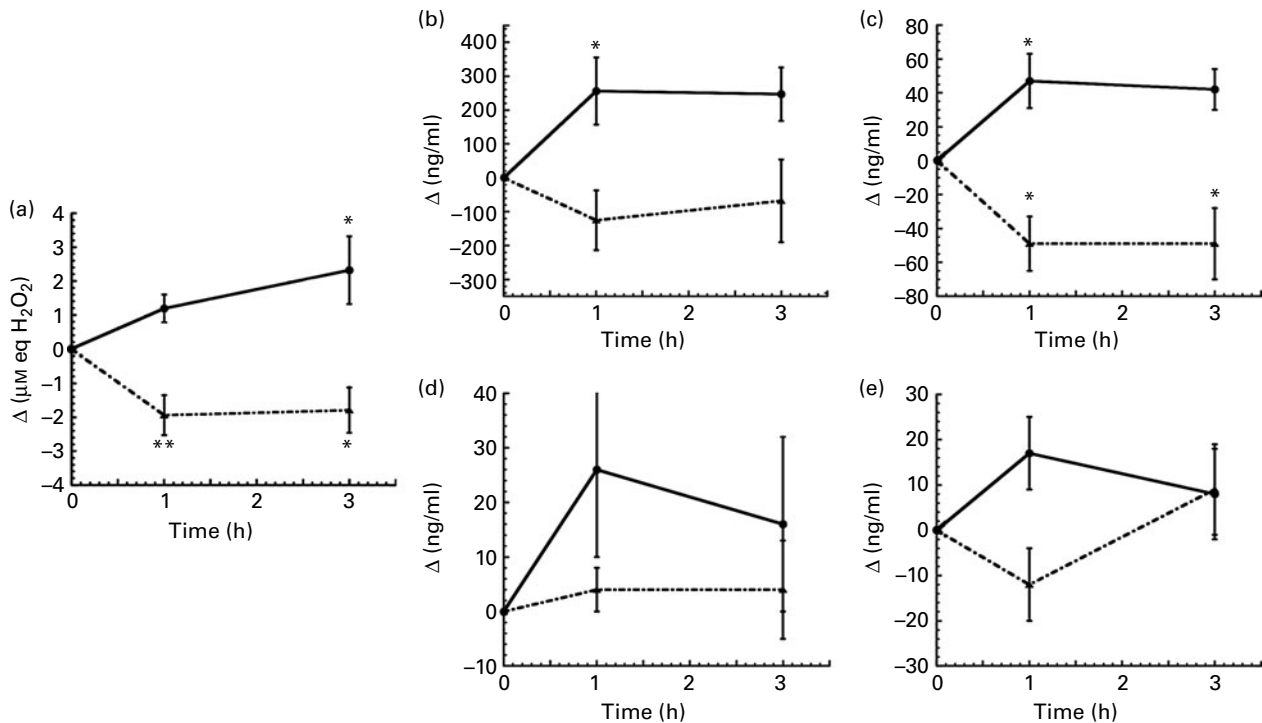
According to the literature, the estimates of the extent to which oxysterols are absorbed vary from 6 to 93%<sup>(28)</sup>. This wide range of results may be due to both the dose and vehicle used to administer the oxysterols<sup>(28)</sup>. Our estimate seems to indicate a complete absorption (105 (SD 29)% of the ingested dose) 1 h after the control meal. We hypothesise, however, that some of the oxysterols present in plasma derive from the oxidation of the cholesterol contained in the meal during the digestive process. Although the formation of oxysterols during digestion cannot be demonstrated by the present study design, several authors provide evidence that lipid oxidation can occur during digestion<sup>(26,27,29)</sup>, and that the presence of antioxidants in the digestive tract can protect from this event<sup>(26,30)</sup>.

A few animal studies have demonstrated that supplementation with antioxidants can prevent the increase in

**Table 1.** Plasma concentration of some metabolic parameters in plasma after a double cheeseburger meal with 300 ml of water or wine (Mean values and standard deviations,  $n$  12)

	Control						Wine					
	0		1 h		3 h		0		1 h		3 h	
	Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD
Total cholesterol (mg/l)	1630	300	1650	260	1610	300	1660	250	1640	270	1660	260
TAG (mg/l)	820	160	820*	240	1070*	330	670	210	970	530	1230*	240
Alcohol (% w/v)	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.08*	0.03	0.05*	0.02

\* Mean values were significantly different from those of homologous time 0:  $P < 0.005$  (by repeated-measures ANOVA, followed by Tukey's test).



**Fig. 1.** Time course of plasma (a) lipid hydroperoxides and oxysterols ((b) 7-ketocholesterol, (c) 7-β-hydroxycholesterol, (d) 5α,6β-epoxycholesterol and (e) 5β,6β-epoxycholesterol) after the administration of the control meal (—) or wine meal (---). Values are means, with standard errors represented by vertical bars (*n* 12). Mean values were significantly different from those of homologous time 0: \* *P*<0.05 and \*\* *P*<0.01 (by repeated-measures ANOVA, followed by Tukey's test).

circulating oxysterols induced by a high-fat diet<sup>(31)</sup>, while the addition of pro-oxidant species to the diet results in a drastic increase in hepatic oxysterols<sup>(32)</sup>. Finally, some human studies have demonstrated that long-term supplementation of antioxidants can reduce the plasma level of oxysterols<sup>(33)</sup>. Thus, the composition of diet (its antioxidant/pro-oxidant balance) has a great influence on the circulating level of oxysterols.

However, in all these studies, the effects of antioxidants on the circulating level of oxysterols have been studied after a chronic supplementation with a high-fat diet. The present study, instead, demonstrates that wine could prevent the acute oxysterol 'toxicity' induced by a single high-fat meal.

It has been demonstrated that wine or wine polyphenols consumption can hinder many harmful postprandial events, such as oxidative stress and endothelial dysfunction. Red wine consumption with the meal reduces the susceptibility to oxidation of postprandial LDL<sup>(3)</sup> and prevents the postprandial increase in plasma lipid hydroperoxides and malondialdehyde<sup>(4)</sup>. A standardised grape product suppresses the meal-induced impairment of vascular endothelial function<sup>(34)</sup>.

In the present study, we have demonstrated for the first time that a glass of wine can prevent the postprandial increase in plasma lipid hydroperoxides and oxysterols after the ingestion of a high-fat, high-cholesterol meal. The peak point seems to correspond to 1 h, but our experimental design (last point 3 h after the meal) cannot indicate the length of the effect; this is a limitation of the present study.

Epidemiological studies have indicated a J-shaped relationship between wine consumption and CVD risk<sup>(35)</sup>. The shape of the curve is the result of the opposite effects of wine/alcohol on the cardiovascular system: 'positive', such as an increase in HDL-cholesterol, anti-thrombotic effects, improved endothelial function, reduced insulin resistance, etc. and 'negative', such as an increase in postprandial TAG level (that is evident also from our data, see TAG in Table 1) and induction of lipid peroxidation by ethanol.

In this view, the postprandial reduction in oxysterols and oxidised lipids could represent a further 'positive' effect of wine. It is well known, in fact, that oxysterols are present in atherosclerotic lesions<sup>(36–38)</sup> and atherogenic lipoprotein<sup>(39)</sup>, and possess several proatherogenic activities, such as cytotoxicity on endothelial and arterial smooth muscle cells, down-regulation of LDL receptors on vascular cell, proinflammatory activities (induction of cytokine release by macrophages and of the expression of adhesion molecule in endothelial cells)<sup>(40–45)</sup>. Finally, several animal studies have shown that oxysterols promote the onset and the development of atherosclerosis<sup>(9,46,47)</sup>.

The results of the present pilot study do not allow explanation of the mechanisms/reactions by which wine counteracts the postprandial increase in circulating oxidised lipids, so that we can just speculate on some possibilities, needing experimental confirmation.

Wine polyphenols and/or alcohol could minimise the postprandial increase in plasma lipid hydroperoxides and cholesterol oxidation products by (1) reducing lipid peroxidation

products or preventing their formation in the digestive tract<sup>(26)</sup>, (2) preventing or delaying fat absorption<sup>(48–50)</sup>, (3) inducing detoxifying enzymes in the gut and liver<sup>(51,52)</sup>, (4) enhancing the cholesterol oxidation product clearance, through the induction of enzymes involved in the cholesterol catabolism towards bile acids<sup>(53,54)</sup> and (5) chemically reducing lipid hydroperoxide and/or oxysterols into the circulation after their absorption.

We studied the effect of wine as a whole, thus we cannot determine which is the wine component (alcohol or polyphenols) responsible for the observed effects and whether other forms of alcoholic beverages could have similar effects. This matter is definitely very interesting, and it should be the object of further investigation. Similarly, it should be important to study how a different ratio of wine:meal oxysterols could affect the wines capacity to cope with the increase in plasma oxysterols.

The present study provides evidence that consumption of wine with a meal could prevent and 'counterattack' the postprandial increase in plasma lipid hydroperoxides and oxysterols, thus protecting the organism from their potential proatherogenic effect. In this view, the controversial effect of a moderate wine consumption on 'health' (different effects *v.* different diseases) could be revised, as the modality of drinking wine (either during or separately from the meal) could represent a decisive factor.

### Acknowledgements

We acknowledge the financial support by grant 'NUME' (DM 3688/7303/08) from the Italian Ministry of Agriculture, Food and Forestry. We thank all the volunteers for their participation. Kariklia Pascucci is acknowledged for her kind support in the daily laboratory work. F. N., A. M., A. R., F. M., R. M. M. and C. S. were responsible for the study design, endpoint assays, data analyses and interpretation, and writing of the manuscript. M. F. assisted in conducting of the experiments. All authors reviewed the manuscript and provided scientific and editorial input. None of the authors had a conflict of interest.

### References

1. Kloner RA & Rezkalla SH (2007) To drink or not to drink? That is the question. *Circulation* **116**, 1306–1317.
2. Covas MI, Gambert P, Fito M, *et al.* (2010) Wine and oxidative stress: up-to-date evidence of the effects of moderate wine consumption on oxidative damage in humans. *Atherosclerosis* **208**, 297–304.
3. Natella F, Ghiselli A, Guidi A, *et al.* (2001) Red wine mitigates the postprandial increase of LDL susceptibility to oxidation. *Free Radic Biol Med* **30**, 1036–1044.
4. Gorelik S, Ligumsky M, Kohen R, *et al.* (2008) A novel function of red wine polyphenols in humans: prevention of absorption of cytotoxic lipid peroxidation products. *FASEB J* **22**, 41–46.
5. Natella F, Fidale M, Tubaro F, *et al.* (2007) Selenium supplementation prevents the increase in atherogenic electro-negative LDL (LDL minus) in the postprandial phase. *Nutr Metab Cardiovasc Dis* **17**, 649–656.

6. Vogel RA, Corretti MC & Plotnick GD (1997) Effect of a single high-fat meal on endothelial function in healthy subjects. *Am J Cardiol* **79**, 350–354.
7. Twickler TB, Dallinga-Thie GM, Visseren FL, *et al.* (2003) Induction of postprandial inflammatory response in adult onset growth hormone deficiency is related to plasma remnant-like particle-cholesterol concentration. *J Clin Endocrinol Metab* **88**, 1228–1233.
8. Leonarduzzi G, Sevanian A, Sottero B, *et al.* (2001) Up-regulation of the fibrogenic cytokine TGF-beta1 by oxysterols: a mechanistic link between cholesterol and atherosclerosis. *FASEB J* **15**, 1619–1621.
9. Staprans I, Pan XM, Rapp JH, *et al.* (1998) Oxidized cholesterol in the diet accelerates the development of aortic atherosclerosis in cholesterol-fed rabbits. *Arterioscler Thromb Vasc Biol* **18**, 977–983.
10. Zieden B, Kaminskas A, Kristenson M, *et al.* (1999) Increased plasma 7 beta-hydroxycholesterol concentrations in a population with a high risk for cardiovascular disease. *Arterioscler Thromb Vasc Biol* **19**, 967–971.
11. Vejux A & Lizard G (2009) Cytotoxic effects of oxysterols associated with human diseases: induction of cell death (apoptosis and/or oncosis), oxidative and inflammatory activities, and phospholipidosis. *Mol Aspects Med* **30**, 153–170.
12. van de Bovenkamp P, Kosmeijer-Schuil TG & Katan MB (1988) Quantification of oxysterols in Dutch foods: egg products and mixed diets. *Lipids* **23**, 1079–1085.
13. Canali R, Ambra R, Stelitano C, *et al.* (2007) A novel model to study the biological effects of red wine at the molecular level. *Br J Nutr* **97**, 1053–1058.
14. Mattivi F, Vrhovsek U, Masuero D, *et al.* (2009) Differences in the amount and structure of extractable skin and seed tannins amongst red grape cultivars. *Aust J Grape Wine Res* **15**, 27–35.
15. Nourooz-Zadeh J (1999) Ferrous ion oxidation in presence of xylenol orange for detection of lipid hydroperoxides in plasma. *Methods Enzymol* **300**, 58–62.
16. Grau A, Codony R, Rafecas M, *et al.* (2000) Lipid hydroperoxide determination in dark chicken meat through a ferrous oxidation-xylenol orange method. *J Agric Food Chem* **48**, 4136–4143.
17. Hahn C, Reichel C & von Bergmann K (1995) Serum concentration of 7 alpha-hydroxycholesterol as an indicator of bile acid synthesis in humans. *J Lipid Res* **36**, 2059–2066.
18. Staprans I, Pan XM, Rapp JH, *et al.* (2003) Oxidized cholesterol in the diet is a source of oxidized lipoproteins in human serum. *J Lipid Res* **44**, 705–715.
19. Guardiola F, Bou R, Boatella J, *et al.* (2004) Analysis of sterol oxidation products in foods. *J AOAC Int* **87**, 441–466.
20. Baggio SR, Miguel AMR & Bragagnolo N (2005) Simultaneous determination of cholesterol oxides, cholesterol and fatty acids in processed turkey meat products. *Food Chem* **89**, 475–484.
21. Rodriguez-Estrada MT, Penazzi G, Caboni MF, *et al.* (1997) Effect of different cooking methods on some lipid and protein components of hamburgers. *Meat Sci* **45**, 365–375.
22. Cameron JR, Skofronick JG & Grant RM (1999) *Physics of the Body*, 2nd ed. pp. 182. Madison, WI: Medical Physics Publishing.
23. Cohn JS (2002) Oxidized fat in the diet, postprandial lipaemia and cardiovascular disease. *Curr Opin Lipidol* **13**, 19–24.
24. Staprans I, Pan XM, Rapp JH, *et al.* (2000) Oxidized cholesterol in the diet accelerates the development of atherosclerosis in LDL receptor- and apolipoprotein E-deficient mice. *Arterioscler Thromb Vasc Biol* **20**, 708–714.



25. Linseisen J & Wolfram G (1998) Absorption of cholesterol oxidation products from ordinary foodstuff in humans. *Ann Nutr Metab* **42**, 221–230.
26. Kanner J & Lapidot T (2001) The stomach as a bioreactor: dietary lipid peroxidation in the gastric fluid and the effects of plant-derived antioxidants. *Free Radic Biol Med* **31**, 1388–1395.
27. Terao J, Ingemansson T, Ioku K, *et al.* (1995) Effects of rat bile-pancreatic juice on Fe<sup>2+</sup> induced peroxidation of phospholipids. *Biosci Biotechnol Biochem* **59**, 55–58.
28. Brown AJ & Jessup W (1999) Oxysterols and atherosclerosis. *Atherosclerosis* **142**, 1–28.
29. Gorelik S, Ligumsky M, Kohen R, *et al.* (2008) The stomach as a 'bioreactor': when red meat meets red wine. *J Agric Food Chem* **56**, 5002–5007.
30. Li Z, Henning SM, Zhang Y, *et al.* (2010) Antioxidant-rich spice added to hamburger meat during cooking results in reduced meat, plasma, and urine malondialdehyde concentrations. *Am J Clin Nutr* **91**, 1180–1184.
31. Ogino Y, Osada K, Nakamura S, *et al.* (2007) Absorption of dietary cholesterol oxidation products and their downstream metabolic effects are reduced by dietary apple polyphenols. *Lipids* **42**, 151–161.
32. Brandsch C, Ringseis R & Eder K (2002) High dietary iron concentrations enhance the formation of cholesterol oxidation products in the liver of adult rats fed salmon oil with minimal effects on antioxidant status. *J Nutr* **132**, 2263–2269.
33. Porkkala-Sarataho E, Salonen JT, Nyyssönen K, *et al.* (2000) Long-term effects of vitamin E, vitamin C, and combined supplementation on urinary 7-hydro-8-oxo-2'-deoxyguanosine, serum cholesterol oxidation products, and oxidation resistance of lipids in nondepleted men. *Arterioscler Thromb Vasc Biol* **20**, 2087–2093.
34. Chaves AA, Joshi MS, Coyle CM, *et al.* (2009) Vasoprotective endothelial effects of a standardized grape product in humans. *Vascul Pharmacol* **50**, 20–26.
35. Klatsky AL (2009) Alcohol and cardiovascular diseases. *Expert Rev Cardiovasc Ther* **7**, 499–506.
36. Hulten LM, Lindmark H, Diczfalussy U, *et al.* (1996) Oxysterols present in atherosclerotic tissue decrease the expression of lipoprotein lipase messenger RNA in human monocyte-derived macrophages. *J Clin Invest* **97**, 461–468.
37. Brown AJ, Leong SL, Dean RT, *et al.* (1997) 7-Hydroperoxycholesterol and its products in oxidized low density lipoprotein and human atherosclerotic plaque. *J Lipid Res* **38**, 1730–1745.
38. Garcia-Cruet S, Carpenter KL, Guardiola F, *et al.* (1999) Oxysterols in cap and core of human advanced atherosclerotic lesions. *Free Radic Res* **30**, 341–350.
39. Babiker A & Diczfalussy U (1998) Transport of side-chain oxidized oxysterols in the human circulation. *Biochim Biophys Acta* **1392**, 333–339.
40. Sevanian A, Berliner J & Peterson H (1991) Uptake, metabolism, and cytotoxicity of isomeric cholesterol-5,6-epoxides in rabbit aortic endothelial cells. *J Lipid Res* **32**, 147–155.
41. Hughes H, Mathews B, Lenz ML, *et al.* (1994) Cytotoxicity of oxidized LDL to porcine aortic smooth muscle cells is associated with the oxysterols 7-ketocholesterol and 7-hydroxycholesterol. *Arterioscler Thromb* **14**, 1177–1185.
42. Aupeix K, Weltin D, Mejia JE, *et al.* (1995) Oxysterol-induced apoptosis in human monocytic cell lines. *Immunobiology* **194**, 415–428.
43. Nishio E & Watanabe Y (1996) Oxysterols induced apoptosis in cultured smooth muscle cells through CPP32 protease activation and bcl-2 protein downregulation. *Biochem Biophys Res Commun* **226**, 928–934.
44. Leonarduzzi G, Sottero B & Poli G (2002) Oxidized products of cholesterol: dietary and metabolic origin, and proatherosclerotic effects (review). *J Nutr Biochem* **13**, 700–710.
45. Poli G, Sottero B, Gargiulo S, *et al.* (2009) Cholesterol oxidation products in the vascular remodeling due to atherosclerosis. *Mol Aspects Med* **30**, 180–189.
46. Vine DF, Mamo CL, Beilin LJ, *et al.* (1998) Dietary oxysterols are incorporated in plasma triglyceride-rich lipoproteins, increase their susceptibility to oxidation and increase aortic cholesterol concentration of rabbits. *J Lipid Res* **39**, 1995–2004.
47. Rong JX, Rangaswamy S, Shen L, *et al.* (1998) Arterial injury by cholesterol oxidation products causes endothelial dysfunction and arterial wall cholesterol accumulation. *Arterioscler Thromb Vasc Biol* **18**, 1885–1894.
48. Pal S, Naissides M & Mamo J (2004) Polyphenolics and fat absorption. *Int J Obes Relat Metab Disord* **28**, 324–326.
49. Ikeda I, Imasato Y, Sasaki E, *et al.* (1992) Tea catechins decrease micellar solubility and intestinal absorption of cholesterol in rats. *Biochim Biophys Acta* **1127**, 141–146.
50. Green PH (1983) Alcohol, nutrition and malabsorption. *Clin Gastroenterol* **12**, 563–574.
51. Moon YJ, Wang X & Morris ME (2006) Dietary flavonoids: effects on xenobiotic and carcinogen metabolism. *Toxicol In vitro* **20**, 187–210.
52. Lu Y, Zhuge J, Wu D, *et al.* (2010) Ethanol-induction of cytochrome P450 2A5: permissive role for CYP2E1. *Drug Metab Dispos*, (Epublication ahead of print version).
53. Yang TT & Koo MW (2000) Chinese green tea lowers cholesterol level through an increase in fecal lipid excretion. *Life Sci* **66**, 411–423.
54. Del Bas JM, Fernandez-Larrea J, Blay M, *et al.* (2005) Grape seed procyanidins improve atherosclerotic risk index and induce liver CYP7A1 and SHP expression in healthy rats. *FASEB J* **19**, 479–481.